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Note

New gas-liquid chromatographic method for biologically important indoleamines

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The presence of enzyme systems producing N- and O-methylated indoleamines in various tissues has prompted the development of procedures aimed at the identification and quantitative estimation of trace amounts of these compounds. Gas-liquid chromatography (GLC) has received increased attention, owing to its high resolving power and sensitivity. In the GLC methods reported the indoleamines were trimethylsilylated¹ in the phenolic group, and or heptafluorobutyrylated², trifluoroacetylated³ or pentafluoropropionylated⁴ in the aminic group.

In connection with our studies on indoleamine-N-methyltransferase⁵, we have developed a procedure for determining tryptamine, N-methyltryptamine, 5-methoxytryptamine and 5-hydroxytryptamine simultaneously by means of flame ionization detection, without derivatization and in a total analysis time of 13 min.

EXPERIMENTAL

Materials

Tryptamine hydrochloride, N-methyltryptamine, 5-hydroxytryptamine oxalate salt and 5-methoxytryptamine hydrochloride were purchased from Sigma, St. Louis, Mo., U.S.A. Other solvents and reagents were obtained from commercial sources.

Apparatus

A Varian Model 3700 gas chromatograph equipped with a hydrogen flame detector and a linear temperature programmer was used. The GLC column (50 cm × 3 mm I.D.) was packed with 5% OV-101 on 100–120 mesh Chromosorb G HP. The injector block was maintained at 200° and the flame ionization detector at 230°. Nitrogen was used as the carrier gas at a constant flow-rate of 20 ml/min. Quantitation of peak area was performed by the peak height × width at half height method.

RESULTS AND DISCUSSION

Fig. 1 shows the chromatogram obtained from 1 μg of a mixture of pure

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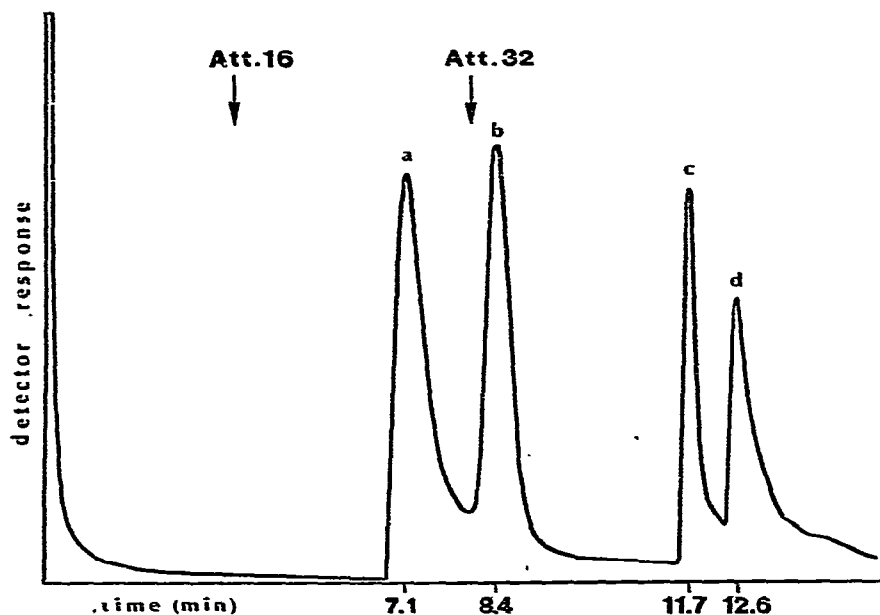


Fig. 1. Gas chromatogram of $1\ \mu\text{g}$ of each indoleamine shown in Table I. Changes in instrument attenuation (Att.) are indicated by vertical arrows.

indoleamines injected in $1\ \mu\text{l}$ of ethyl acetate. In the analysis the oven was maintained at 140° for 7 min, and temperature programming was carried out from 140° to 230° at 15° per min. Under these conditions the peaks are symmetrical and well resolved, thus allowing their complete integration. Fig. 2 shows the standard calibration curves for the indoleamines analysed. Response, as measured by peak area multiplied by attenuation in the gas-liquid chromatogram of indoleamines, was found to be linear,

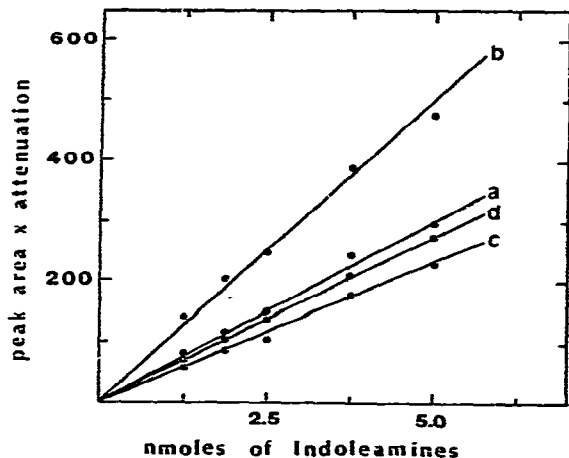


Fig. 2. Linearity plot. The y -axis is the GLC chromatogram peak area multiplied by the instrument attenuation; the x -axis is the concentration of indoleamines in nmoles.

while the emergence times were reproducible to 0.1 min or better. The lower detection limit, for the identification and quantitation was 5 ng. The retention data are given in Table I.

TABLE I
RETENTION DATA FOR STANDARDS

<i>Compound</i>	<i>Emergence time (min)</i>	<i>Peak area × attenuation* (cm²)</i>
(a) Tryptamine	7.1	294
(b) N-Methyltryptamine	8.4	458
(c) 5-Methoxytryptamine	11.7	226
(d) 5-Hydroxytryptamine	12.6	274

* Data correspond to 1 μ g of indoleamine.

This technique is very selective and sensitive for the identification of products of reactions catalysed by hydroxyindole-O-methyltransferase and indoleamine-N-methyltransferase, using 5-hydroxytryptamine and tryptamine respectively as substrates. It is not necessary to use derivatization for the identification and quantitation of 5-methoxytryptamine and N-methyltryptamine. We are presently examining the indoleamine content in various tissues from different animal species. These results will be reported elsewhere.

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